**Effect of Bisphenol-A on Serum Biochemistry and Liver Function in the Freshwater Fish, *Catla catla***

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**ABSTRACT**

Bisphenol-A (BPA) is an anthropogenic chemical mimicking 17β estradiol actions. It is used as a monomer for plastic production. In the present study, adult female (age around 18 months) *Catla catla*, a widespread cyprinid fish in the Pakistani rivers, were exposed to increasing concentrations (10, 100, 1000 µg/l) of BPA for 2-weeks. Classical toxicological endpoints like liver enzymes and alterations in liver histology were investigated. In addition, the effect of BPA on hepatosomatic index (HSI) and vitellogenin (vtg) mRNA expression was also examined. A concentration-dependent increase was recorded in serum liver enzymes like aspartic aminotransferase (AST) and alanine aminotransferase (ALT) compared to control. The biomarkers for kidney function like serum creatinine level and uric acid also significantly increased in fish exposed to BPA. HSI did not change in groups exposed to low concentrations of BPA (10 and 100 µg/l); however, a significant increase as compared to control was observed in female fish exposed to 1000 µg/l BPA. Hepatic vtg mRNA increased with increase in exposure concentration but only the highest concentration (1000µg/l) was capable of inducing a significant vtg expression. Present study showed that bisphenol-A has far reaching detrimental effects on fish health by altering metabolic profile of liver and kidney and induced significant histopathological changes in the liver. Moreover, it also exerted environmental estrogenic effects by inducing vitellogenin, similar to natural estrogens. The results presented here are similar to earlier studies reporting estrogenic actions of BPA.

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**INTRODUCTION**

Aquatic ecosystem receives a large number of pollutants that pose major threats to the aquatic fauna during their developmental and adult life stages. Such anthropogenic chemicals are major problem around the globe and toxicity of anthropogenic chemicals is usually dependent on their concentrations in the environment, persistent nature and bioaccumulation/bioavailability (Sayed et al., 2011).

Bisphenol-A (BPA) is a xenobiotic and solely manmade chemical. BPA is a plastic monomer used for the synthesis of polycarbonate plastics, epoxy resins and thermal papers. It is also used as a precursor of flame retardant (Corrales et al., 2015) and as a constituent of dental sealant (Suzuki et al., 2000). To cope the rising demand of plastic and plastic products, large volume of BPA is produced worldwide (Corrales et al., 2015). Production of BPA in large quantity resulted in subsequent release in the environment especially aquatic environment.

BPA enters into the water bodies through manufacturing plants and effluent discharge. BPA is also released during transport and processing, degradation of plastic and PVC pipes also result in BPA release (Flint et al., 2012). BPA gained much attention in the recent years because it has the affinity for estrogen receptors (ERα and β) present on cell and nuclear membrane and with peroxisome proliferator receptors (PPRs) (Corrales et al., 2015). BPA also interact with thyroid and androgen receptor and alter the titer of androgens and thyroid hormones in the body (Faheem et al., 2017b). In aquatic environment, BPA is usually degraded in 0.5-6 days (Mihaich et al., 2012). Because of its unremitting release...
Biochemical analysis: Serum biochemical parameters e.g. aspartic amino-transferase (AST), alanine amino-transferase (ALT), creatinine (Cr) and uric acid were determined by commercially available kits (Randox reagents).

Histological examinations: The tissue samples preserved in formalin were dehydrated by passing through ascending concentrations of alcohol. After clearing with xylene, the liver tissue were impregnated with wax. Tissue sections were cut (5µ thick). Hematoxylin and eosin (H & E) stains were used to stain the tissue. After staining, slides were examined carefully and photographs were taken with high resolution camera fitted on a microscope (Leica, Japan).

Real time qPCR: Vitellogenin expression in liver was studied by real time qPCR. One microgram of total RNA from liver tissue was reverse transcribed using oligo dT primers. Real-time qPCR was performed using CFX 96 (Bio-Rad) with Syber Green fluorescent label. Geometric mean of three most stable reference genes (18s, gapdh, tbp) was used as reference control following Faheem et al. (2018a). Primers used in gene expression study are listed in Table 1. Melt curve analysis was performed after amplification to ensure amplification specificity. Each sample was run in duplicate and cycle threshold value generated by Software (CFX Manager, Version 3.1). 2–ΔΔct method was used to calculate change in transcript abundance (Livak and Schmittgen, 2001).

Statistical analysis: All data of biochemical and gene expression studies are expressed as means ± S.E.M. One way analysis of variance (ANOVA) and Tukey’s post hoc test was used to determine the statistical difference among means. All the statistical analysis was performed on SPSS (IBM, Version 20). P value less than 0.05 was considered as significant.

RESULTS

No significant change in hepato-somatic index (HSI) was observed in female Catla catla exposed to 10 and 100 µg/l BPA. However, fish exposed to highest concentration (1000 µg/l) of BPA had significantly higher HSI as compared to control group (Fig. 1).

Histological examination of adult female fish liver exposed to increasing concentrations of BPA is given in Fig. 2. Hepatic tissue of fish from control group showed normal architecture. Central vein was lined with epithelial cells and hepatocytes were arranged in cords. Control group hepatocytes had centrally located nucleus with no vacuoles. Similar picture of hepatocytes was observed in the livers of fish exposed to low concentration (10 µg/l) of BPA. Few vacuoles were seen and shrinkage of pancreatic cells (the fish has heptopancreas) were observed in fish liver exposed to 100 µg/l BPA. Major histopathological alterations were only observed in hepatocytes of fish exposed to 1000 µg/l BPA. These alterations involved loss of normal architecture of hepatocytes, shrinkage of pancreatic cells, lipid vacuolization, hemorrhage and necrosis.
Fig. 1: Hepato-somatic index (HSI) of adult female Catla catla exposed to 10, 100 and 1000 µg/l BPA for 14 days. Data are expressed as mean ± SEM. n=3. Different letters indicate significant differences among groups, P<0.05.

Fig. 2: Liver tissue from adult female Catla catla exposed to graded concentration of BPA (10, 100 and 1000 µg/l) for 14 days. Pancreatic cells (Pc); Hepatocytes with central nucleus (H); Sinusoids (S); Supporting tissue (St); Hemolysis (He); Shrinkage of pancreatic cells (PcS); Vacuolization (V); Necrosis (N). H & E stain, 400X.

Fig. 3: Serum level of alanine amino transferase (ALT) and aspartic amino transferase (AST) in female Catla catla exposed to 10, 100 and 1000 µg/l BPA for 14 days. Data are expressed as mean ± SEM. n=3. Different letters indicate significant differences among groups. P<0.05.

An increase in serum ALT and AST (Fig. 3) was recorded but the increase was not significant (P>0.05). A statistically significant increase (P<0.05) in kidney function parameters was recorded after exposure to increasing concentrations of BPA for 14 days. Level of serum creatinine increased significantly in 100 and 1000 µg/l BPA groups (Fig. 4) while uric acid increased only in 1000 µg/l BPA group (Fig. 5).

Table 1: Primer sequences, amplicon lengths and annealing temperature of selected genes

<table>
<thead>
<tr>
<th>Genes</th>
<th>Primer sequence 5’ to 3’</th>
<th>Amplicon size</th>
<th>Annealing temperature (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>gapdh</td>
<td>ATCA-CAGGCCACGCAGAAGACC</td>
<td>126</td>
<td>60</td>
</tr>
<tr>
<td></td>
<td>CAGGAAGCTTTGCGACACG</td>
<td></td>
<td></td>
</tr>
<tr>
<td>l8S</td>
<td>CGGTGAAACCTTGAGACTCT</td>
<td>189</td>
<td>60</td>
</tr>
<tr>
<td></td>
<td>CTTGGAATGTTGAGCCGTTT</td>
<td></td>
<td></td>
</tr>
<tr>
<td>tbp</td>
<td>AACAGCTGGCCCTCTGGA</td>
<td>213</td>
<td>60</td>
</tr>
<tr>
<td></td>
<td>TCCAGGAGGGACAGCTGTT</td>
<td></td>
<td></td>
</tr>
<tr>
<td>vtg</td>
<td>GGTTGCTCCAGACCTTGGAG</td>
<td>180</td>
<td>60</td>
</tr>
<tr>
<td></td>
<td>GCAAGGCTCCACCTTGAG</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Fig. 4: Serum level of uric acid in female Catla catla exposed to 10, 100 and 1000 µg/l BPA for 14 days. Data are expressed as the mean ± SEM. n=3. Different letters indicate significant difference among groups. P<0.05.

Fig. 5: Serum level of creatinine in female Catla catla exposed to 10, 100 and 1000 µg/l BPA for 14 days. Data are expressed as the mean ± SEM. n=3. Different letters indicate significant difference among groups. P<0.05.

Fig. 6: Relative gene expression of the biomarker gene vitellogenin (vtg) normalized to different reference genes and mean of three selected genes (gapdh, l8S, tbp), in liver of adult female Catla catla after 14 days of exposure to graded concentrations of BPA. Data are expressed as the mean fold change ± SEM. n=3. Different letters indicate significant differences among groups. P<0.05.
Vitelligenin mRNA expression did not change in group exposed to 10µg/l but with an increase in concentration, vtg mRNA increased. A significant (P<0.05) induction of vtg mRNA was only observed in group exposed to 1000µg/l/BPA (Fig. 6).

**DISCUSSION**

Vital organs of fish e.g. liver, kidney, gills and brain are important target of any toxicant. Various studies showed that alteration in physiological and blood biochemical parameters are caused by contaminants in the surrounding environment (Ololade and Oginni, 2010; Sayed and Hamed 2017). Study of serum biochemistry provides a valid indication of damage by pollutant and are mirror image of the environmental pollution and are useful for detection of tissue pathophysiological status (Sayed and Hamed 2017; Abdel-Tawwab and Hamed, 2018).

Increased level of serum ALT and AST indicate liver damage (Bhattacharya et al., 2008) and have been used as toxicopathological biomarker (Coppo et al., 2016). Similar increase in level of serum AST and ALT was reported in heavy metal exposed fish (Mekkawy et al., 2011) and other endocrine disrupting chemicals (Bhattacharya et al., 2008). In the present study, exposure of increasing concentrations of BPA to adult female *Catla catla* caused non-significant increase in activity of both enzymes. Similar non-significant increase in AST and ALT was reported in *C. gariepinus* exposed to diazinon (Adedeji et al., 2009). Significant increase in hepatic enzymes (ALT, ALP, AST) was recorded in *Oreochromis niloticus* exposed to 1.64 µg/l BPA for 6 weeks (Abdel-Tawwab and Hamed, 2018). This difference in hepatic enzyme level may be due to different concentrations and exposure time. In our study acute exposure of BPA was used while Abdel-Tawwab and Hamed (2018) used chronic exposure in their study.

Serum creatinine and uric acid are important indicators of renal health and kidney function and biomarkers for muscle and purine metabolism (Hamed and Tawwab, 2017). Glomerulus damage, impairment metabolism of carbohydrates and increased muscle tissue catabolism therefore may cause increased creatinine level in blood (Hadi et al., 2009). A significant increase was recorded in serum creatinine level of fish exposed to 100 and 1000µg/l BPA while uric acid level significantly increased only in fish exposed to 1000 µg/l BPA. Significant increase in creatinine and uric acid may indicate that BPA affect muscle and purine (nucleic acid) metabolism. This increase may also be due to the damage of renal tubules. Degeneration and necrosis of glomerulus and decrease in hematopoietic tissue in the same fish species after BPA exposure was reported by Faheem et al. (2016). This decrease in hematopoietic tissue may be a cause of increase in serum uric acid.

Histopathological evaluation of key organs is an important tool to evaluate the effect of a toxicant on fish health. Liver is the main organ of detoxification, any alteration in the liver tissue may lead to compromised functions in fish. Exposure of BPA to female *Catla catla* resulted in degenerative changes in hepatocytes. Similar histopathological alteration was recorded in *Clarias gariepinus* liver exposed to nonylphenol for 15 days (Sayed et al., 2012). Alteration in liver histology includes change in normal structure, vacuolation of hepatocytes and dilatation of sinusoids. Structural protein degeneration may be the reason of sinusoid dilations after BPA exposure. Vacuolation of hepatocytes is non-specific response of fish after exposure to a toxicant (Roberts, 1978), hepatocyte vacuolation observed here may be due to lipid accumulation. Juvenile *Sparus aurata* exposed to BPA had severe lipid accumulation and degeneration of liver parenchyma and hepatocytes (Maradonna et al., 2014). Exposure of 1000µg/l BPA to *Oreochromis mossambicus* for 10 and 20 days resulted in large amount of hepatocyte vacuolization (Chitra and Maiby, 2014).

Vitellogenin (vtg) is the precursor of egg yolk and used as a biomarker for estrogenic endocrine disruption (Matozzo et al., 2008). Vitellogenin is synthesized in female fish under the influence of endogenous estrogenic hormones. Increased mRNA level of vitellogenin or presence of vitellogenin protein in serum of male and juvenile fish or abnormal level of vitellogenin in female fish, out of reproductive season, is the sign of endocrine disruption. Increased level of hepatic vtg mRNA in female fish suggests BPA has estrogenic activity in *Catla catla*. Results reported here are in accordance with the recent study where administration of 1-100µg/g BPA to immature yellowfin seabream resulted in increased hepatosomatic index and serum vitellogenin (Negintaji et al., 2018). Increased vitellogenin level was also reported in *Cyprinus carpio* exposed to similar chemical and experimental conditions (Mandich et al., 2007). Significant induction of vitellogenin after BPA exposure was earlier reported in different species of fish (Sohoni et al., 2001; Ishibashi et al., 2005; Lee et al., 2007; Mandich et al., 2007; Faheem et al., 2017c; Negintaji et al., 2018). All these studies reported that BPA is capable of vtg induction; however difference among studies can be due to difference in fish species used as model, species-specific estrogen receptor binding, water temperature and exposure time (Craig et al., 2007; Faheem et al., 2017c).

Present study indicate that bisphenol-A at environmentally relevant concentration is capable of inducing vtg mRNA and damage vital organs of fish leading to altered level of enzymes that can cause potential harm to fish health and reproduction. If such fish with heavy load of BPA are consumed regularly by humans also can create similar problems to their health.

Authors contribution: The study was supervised by KPL. All experiments were performed by MF and real time was performed by MF and SK. MF prepared the manuscript.

REFERENCES


