EFFECTS OF GOSSYPOL ON SEMEN CHARACTERISTICS IN TEDDY MALE GOATS

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ABSTRACT

In the present study, the effects of gossypol on semen characteristics in Teddy male goats were studied. Nine Teddy male goats were randomly divided into three equal groups named A, B and C. Animals in all groups were fed concentrated rations without cottonseed cakes (CSC) at the rate of 3% of their liveweight for a period of 30 days and it was named as pre-treatment period. Just after the completion of this period, animals in group A were fed control ration (without gossypol), those in group B were fed ration which contained unboiled CSC as a source of free and bound gossypol, while animals in group C were given ration containing CSC boiled at 100°C for 1 hour as a source of bound gossypol. These experimental rations were fed to animals of respective groups at the rate of 3% of their liveweight for a period of 90 days and it was named as treatment period. Feeding of ration containing gossypol to Teddy male goats did not affect the colour, volume, mass activity, sperm concentration, percentage of dead spermatooza, liveability and absolute index of liveability of spermatooza at 37°C. However, it affected significantly (P<0.05) the pH, per cent motility of spermatooza and percentage of morphologically normal spermatooza. The Teddy male goats fed rations containing a combination of free and bound gossypol showed a significant (P<0.05) increase in the pH and a decrease in motility of spermatooza which was statistically lower than those fed control diet or diet containing bound gossypol. It was concluded that rations containing a combination of free and bound gossypol (unboiled CSC) or bound gossypol only (boiled CSC) adversely affected the semen quality of Teddy male goats in terms of sperm motility and morphologically normal spermatooza in ejaculates.

Key Words: Gossypol, cottonseeds cakes, Teddy male goats, semen characteristics.

INTRODUCTION

Goat breeding, a very basic form of livestock production, is practiced worldwide and makes an important contribution to overall global agricultural production. Goat breeding plays a vital role in the improvement of economy of many people, particularly those in the more primitive areas of sparse vegetation where rearing of other livestock is difficult, the goat is an important provider of milk, meat, and fibers such as mohair, cashmere, and cashgora. Goat farming appears to be an attractive proposition for a small farm, since the milk yield of a good dairy goat is at least 50 per cent greater than that of a good dairy cow, calculated on a weight for weight comparison (Mowlem, 1985).

Gossypol is a yellow pigment, found in various parts of the cotton plants, including seeds, of the genus Gossypium (Adams et al., 1960). Chemically, it is 1, 1', 6, 6', 7, 7'-hexahydroxy-5, 5'-disisopropyl-3, 3'-dimethyl (2', 2'-binaphthalene)-8,8'-dicarboxaldehydehyd with the empirical/molecular formula of C_{22}H_{38}O_{8} (Abou-Donia, 1976) and a molecular weight of 518.54 (Adams et al., 1960). Gossypol is known to cause toxicity in monogastric (Haschek et al., 1989) as well as ruminant animals (Holmberg and Kutches, 1991). This compound has been shown to exert antifertility effects in males (Randel et al., 1992) and females (Gu et al., 1990). It has direct damaging effects on the testes and the developing germ cells (Zahid et al., 2002). Cottonseed cakes contain 0.28 ± 0.02 per cent free and 1.44 ± 0.04 per cent total gossypol (Zahid, 2002).

Cottonseeds and their by-products, e.g., cottonseeds cakes (CSC) and cottonseed meals (CM), have been used extensively as protein supplements in dairy animals' rations to increase fat and milk production in these animals (Ahmad, 1993). As a cheaper but rich source of high quality protein for supplementing animal diets, the use of cottonseed products has been continued over the last many decades. This indiscriminate use of
cottonseed or their by-products can adversely affect the reproductive performance of dairy animals.

However, there is little information in the literature on the effects of gossypol on the semen quality of Teddy male goats. Therefore, the present study was carried out to determine the effects of gossypol on semen characteristics in Teddy male goats.

**MATERIALS AND METHODS**

In the present study, nine adult healthy (10-11 months of age) Teddy male goats were used. These were randomly divided into three groups A, B and C, with three animals in each group. Male goats in all groups were kept under the same managerial and environmental conditions. In order to let the animals adjust to the new environment and adopt themselves to concentrate feeding, all the experimental animals were fed concentrated ration other than cottonseed cakes at the rate of 3% of their liveweight for a period of 30 days, in addition to good quality chaffed seasonal green fodder and clean water ad libitum. This period was named as pre-treatment period. Just after the completion of this period, animals in group A were fed control ration without cottonseed cakes (without gossypol), those in group B were fed ration consisting of unboiled cottonseed cakes which contained both free and bound gossypol (620 and 1750 ppm, respectively), while animals in group C were given ration containing cottonseed cakes boiled at 100°C for 1 hour which contained bound gossypol only (2370 ppm). These rations were fed to animals of respective groups at the rate of 3% of their liveweight for a period of 90 days, in addition to good quality chaffed seasonal green fodder and clean water ad libitum. This period was named as treatment period. During this period, semen samples were collected on weekly intervals by using artificial vagina. On each collection day two consecutive ejaculates were taken from each animal in all experimental groups. Immediately after collection, the semen collecting tubes were taken to the laboratory and placed in a water bath at 37°C. First and second ejaculates were pooled and each pooled sample was evaluated for its physical characteristics, i.e. colour, volume, pH, mass activity, percent motility of spermatozoa, sperm concentration, dead spermatozoa, per cent morphologically abnormal spermatozoa, liveability of spermatozoa at 37°C and absolute index of liveability of spermatozoa, using the standard procedures for evaluation of semen. To measure the volume of semen, two consecutive ejaculates were averaged.

Statistical analysis was performed using analysis of variance technique (Steel and Torrie, 1984) under completely randomised design. For this purpose, general linear model procedure under SAS computer programme (SAS, 1990) was adopted. Least significant difference test was used for comparison of means (Steel and Torrie, 1984).

**RESULTS AND DISCUSSION**

At the completion of treatment period, the mean (± SE) values for characteristics of semen in male goats of groups A, B and C are presented in Table 1. The highest values for semen volume (0.79 ± 0.04 ml) were noted in male goats of control group, while the lowest (0.68 ± 0.04 ml) in animals of group B fed diet containing unboiled cottonseed cakes. Statistical analysis of the data revealed that the difference was non-significant (P>0.05) among all the groups. Similar trend was observed for mass activity (scores), motility of spermatozoa (%), liveability of spermatozoa (hours) and absolute index of liveability of spermatozoa at 37°C. The pH of ejaculates in animals of group B was greater (6.73 ± 0.01) than animals of groups A and C for which these values were 6.69 ± 0.01 and 6.70 ± 0.01, respectively. Statistical analysis of the data revealed that the difference was significant (P<0.05) between groups A and B, and B and C while it was non-significant (P>0.05) between groups A and C.

The values for sperm concentration were higher in group A (3.09 ± 0.01 x 10⁹/ml) while lower values (3.08 ± 0.01 x 10⁹/ml) were observed in groups B and C. However, the difference was non-significant (P>0.05) among all the groups. The values for dead spermatozoa were highest in group B (17.80 ± 1.09%) while lowest (15.90 ± 1.09%) in group A. However, statistically all the groups differed non-significantly (P>0.05). The values for morphologically abnormal spermatozoa were highest in group B (7.62 ± 0.22%) while lowest (3.69 ± 0.22%) in group A. Statistical analysis of the data revealed that the difference was significant (P<0.05) among all the groups (Table 1).

Information on the effects of gossypol feeding on semen characteristics in Teddy male goats and other breeds of goats is scanty. However, Jaimudeen et al. (1982) and Akhtar (1997) observed no change in the colour or mass activity of ejaculates of buffalo bulls fed rations containing gossypol. The results of the present study are in line with the observations of these workers. The results of the present study regarding the ejaculatory volume are in agreement with the findings of Smith et al. (1991) and Akhtar (1997), who reported no change in volume of semen of Holstein and Nili-Ravi buffalo bulls, respectively, fed on rations containing gossypol. Similarly, the percentage of dead
Table 1: Effects of gossypol on semen characteristics in Teddy male goats

<table>
<thead>
<tr>
<th>Semen characteristics</th>
<th>Group A</th>
<th>Group B</th>
<th>Group C</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ejaculatory volume (ml)</td>
<td>0.79 ± 0.04</td>
<td>0.68 ± 0.04</td>
<td>0.76 ± 0.04</td>
</tr>
<tr>
<td>The pH of ejaculates</td>
<td>6.69 ± 0.01²</td>
<td>6.73 ± 0.01²</td>
<td>6.70 ± 0.01¹</td>
</tr>
<tr>
<td>Mass activity (Scores)</td>
<td>4.01 ± 0.04</td>
<td>3.95 ± 0.04</td>
<td>3.99 ± 0.04²</td>
</tr>
<tr>
<td>Motility of spermatozoa (%)</td>
<td>79.66 ± 0.40²</td>
<td>77.36 ± 0.40²</td>
<td>79.70 ± 0.40²</td>
</tr>
<tr>
<td>Sperm concentration (x 10⁹/ml)</td>
<td>3.09 ± 0.01</td>
<td>3.08 ± 0.01</td>
<td>3.08 ± 0.01²</td>
</tr>
<tr>
<td>Dead spermatozoa (%)</td>
<td>15.90 ± 1.09</td>
<td>17.80 ± 1.09</td>
<td>15.93 ± 1.09²</td>
</tr>
<tr>
<td>Morphologically abnormal</td>
<td>3.69 ± 0.22²</td>
<td>7.62 ± 0.22²</td>
<td>5.88 ± 0.22²</td>
</tr>
<tr>
<td>Spermatozoa (%)</td>
<td>7.67 ± 0.75</td>
<td>6.11 ± 0.75</td>
<td>6.78 ± 0.75²</td>
</tr>
<tr>
<td>Liveability of spermatozoa at 37°C (hours)</td>
<td>149.67 ± 13.21</td>
<td>115.33 ± 13.21</td>
<td>126.67 ± 13.21</td>
</tr>
</tbody>
</table>

Group A = Animals were fed ration without gossypol
Group B = Animals were fed ration containing unboiled cottonseed cakes (combination of free and bound gossypol)
Group C = Animals were fed ration containing boiled cottonseed cakes at 100°C for 1 hour (bound gossypol only)

Values bearing different superscripts in the same row differ significantly (P<0.05).

Spermatozoa, liveability and absolute index of liveability of spermatozoa at 37°C and sperm concentration were not affected by feeding rations containing gossypol. Akhtar (1997) also reported non-significant (P>0.05) differences in these parameters of semen collected from Nili-Ravi buffalo bulls fed rations with or without gossypol.

The feeding of ration containing gossypol affected the pH, motility of spermatozoa and percentage of morphologically abnormal spermatozoa. Teddy male goats fed rations containing a combination of free and bound gossypol showed an increase in the pH. However, the pH of semen collected from all these groups remained within the range reported by Mann (1981). The percentage of motility of spermatozoa in animals fed a combination of free and bound gossypol was decreased and statistically it was lower than those fed control diet or ration containing bound gossypol. These results are supported by the findings on semen of man (Hong et al., 1989), rats (Swan et al., 1990) and Brahman bulls (Chenoweth et al., 1994), where a deleterious effect of free gossypol feeding on sperm motility was reported.

In the present study, feeding of rations containing a combination of free and bound gossypol or bound gossypol only, adversely affected the morphologically abnormal spermatozoa. Similarly, Arshami and Ruttle (1989) reported higher percentage of abnormal spermatozoa in the ejaculates from rams fed rations containing 14 to 17 per cent cottonseeds (gossypol) than those fed control ration.

Morphology of spermatozoa plays an important role in their fertilizing ability. An increased number of morphologically abnormal spermatozoa in an ejaculate is associated with a decrease in fertility (Lagerlof, 1934). Infertility is a reflection of lesions in the testes and/or of the excurrent duct system. Hikam and Hoffer (1987) reported reduced spermatogenesis accompanied by increased abnormal sperms in rats after feeding gossypol. An increased number of morphologically abnormal spermatozoa in ejaculates has been reported following feeding ration containing gossypol to rats (Swan et al., 1990) and cocks (Mohn et al., 1989). The results of the present study are supported by the findings of these workers as the feeding of ration containing gossypol showed a significant increase in the pH and a decrease in motility of spermatozoa which was statistically lower than those fed control diet or diet containing bound gossypol.

Based on the findings of the present study, it can be concluded that rations containing a combination of free and bound gossypol (unboiled CSC) or bound gossypol only (boiled CSC) adversely affected the semen quality of Teddy male goats in terms of pH, sperm motility and morphologically abnormal spermatozoa in ejaculates. Thus, gossypol seems to be a potential antifertility agent, as increased number of abnormal spermatozoa, increase in pH and decrease in sperm motility in an ejaculate are associated with a decrease in fertility.
REFERENCES


