

Pakistan Veterinary Journal

ISSN: 0253-8318 (PRINT), 2074-7764 (ONLINE) DOI: 10.29261/pakvetj/2018.036

RESEARCH ARTICLE

Polyherbal Formulation Ameliorates Diabetes Mellitus in Alloxan-Induced Diabetic Rats: Involvement of Pancreatic Genes Expression

Asra Iftikhar, Bilal Aslam*, Faqir Muhammad and Tanweer Khaliq

Institute of Pharmacy, Physiology and Pharmacology, University of Agriculture, Faisalabad, Pakistan *Corresponding author: cba933@gmail.com

ARTICLE HISTORY (17-338)

Received: December 26, 2017 Revised: January 08, 2018 Accepted: January 12, 2018 Published online: March 28, 2018 Key words: Alloxan monohydrate Diabetes Gene expression Ins-1 mRNA Pdx-1

ABSTRACT

Medicinal plants and herbal formulations have been used as folk medicines for the treatment of hyperlipidemia, diabetes and related metabolic disorders in Asian counties. Current study was designed to evaluate the therapeutic potential of polyherbal formulation consisting of Swertia chirata (Chirata), Artemisia absinthium (Afsanteen boti), Caesalpinia bonduc (kranjwa magz), Bunium persicum (Kala zeera), Gymnema sylvestra (Ghurmar boti), Citrullus colocynthis (Kortumma), Sphaeranthus indicus (Mundi boti) and Cuminum cyminum (Safeed zeera) in alloxan induced diabetic rats. Rats were grouped into 6 groups and induced diabetes using 150mg/kg alloxan monohydrate except control group. Group I served as control, group II was diabetic group and group III was considered +ve control (glibenclamide treated). Group IV, V and VI were treated with 200, 400 and 600mg/kg of polyherbal formulation upto 8 weeks of experiment. Sampling for biochemical parameters and gene expression analysis was done at 8th week of experiment. Results have shown that polyherbal formulation produced anti-diabetic and antihyperlipidemic effects by reducing serum glucose, glycosylated hemoglobin, total cholesterol, triglycerides, LDL, VLDL levels and increasing serum insulin, glucokinase and HDL levels. Gene expression analysis has revealed that polyherbal formulation upregulated the mRNA expression of Ins-1 and Pdx-1 genes in alloxanized hyperglycemic rats. All these properties of polyherbal formulation make it a potential candidate for the treatment of diabetes. However, further research is required to elucidate its mechanism of action.

©2018 PVJ. All rights reserved

To Cite This Article: Iftikhar A, Aslam B, Muhammad F and Khaliq T, 2018. Polyherbal formulation ameliorates diabetes mellitus in alloxan-induced diabetic rats: involvement of pancreatic genes expression. Pak Vet J, 38(3): 261-265. <u>http://dx.doi.org/10.29261/pakvetj/2018.036</u>

INTRODUCTION

Diabetes mellitus is a metabolic disorder characterized by high blood glucose level because the beta (β) cells of pancreas do not produce adequate insulin, or insulin does not produce response at particular receptors (Riserus et al., 2009). Diabetes mellitus is considered as third greatest "killer" after cancer and cardiovascular diseases (Hatware and Annapurna, 2014). The number of diabetic patients is increasing day by day in both rich and developing countries. According to statistics, 5% of all casualties in the world are caused by diabetes, a number which is expected to rise by 50% in next 10 years (Piya et al., 2010). Chronic hyperglycemic effects of diabetes account for various types of abnormalities such as failure and dysfunction of the eyes, nerves, heart, kidneys and blood vessels (Si et al., 2011). As there is no specific

remedy for diabetes, so the most important goal in the cure of all types of diabetes is preservation and even potential regeneration of pancreatic β - cells.

From past three decades, regardless of the huge advancement made in the treatment of diabetes, the patients are still suffering from complications (Dey *et al.*, 2002). Despite significant improvement made by synthetic drugs in the management and treatment of diabetes mellitus, the search for new natural anti-diabetic agents is going on due to least side effects associated with plants. According to an estimate made by world health organization (WHO), 80% of people particularly in developing countries are using natural medicines for treatment of different diseases (Almalki *et al.*, 2016). Many plants are used in folk medicines due to their antihyperglycemic potential. To date, more than 800 species of plants have been investigated and their antidiabetic effects are reported (Mannan *et al.*, 2014).

A number of medicinal plants have been reported for therapeutic properties (Hossen et al., 2016; Liagat et al., 2016; Abbas et al., 2017a; Hassan et al., 2017; Tahir et al., 2017) mainly due to the presence of different complex substances known as secondary metabolites such as flavonoids, polyphenols and alkaloids (El-Abhar and Schaalan, 2014; Abbas et al., 2017b). Scientific research studies have also reported the use of polyherbal formulations for the treatment of diabetes. Polyherbal formulation considerably alters the pattern of glucose tolerance, lipids profile and other biochemical parameters in diabetes mellitus (Ghorbani, 2014). It is believed that herbal formulations containing multiple plant products produce synergistic antidiabetic effects and could enhance the desired actions. The polyherbalism improves the therapeutic efficiency of formulation and reduces unwanted adverse effects (Paoli et al., 2015). So the present research work was designed to assess the potential of polyherbal formulation consisting of Artemisia absinthium (Afsanteen boti), Sphaeranthus indicus (Mundi boti), Swertia chirata (Chirata), Gymnema sylvestra (Ghurmar boti), Caesalpinia bonduc (kranjwa magz), Bunium persicum (Kala zeera), Cuminum cyminum (Safeed zeera) and Citrullus colocynthis (Kortumma) on glycemic levels, lipid profile and different genes involved in β cells regeneration and insulin secretion from β cells in alloxanized hyperglycemic rats.

MATERIALS AND METHODS

Preparation of polyherbal formulation: For polyherbal formulation preparation, leaves of Artemisia absinthium, Gymnema sylvestra, Sphaeranthus indicus and Swertia chirata, fruits of Caesalpinia bonduc and Citrullus colocynthis, seeds of Bunium persicum and Cuminum cyminum were used. Samples were verified for taxonomy from Department of Botany and a voucher specimen of each plant was deposited in the herbarium of University of Agriculture, Faisalabad, (voucher specimen numbers: (Gymnema sylvestra:21144), (Artemisia absinthium: 21145), (Sphaeranthus indicus:21146), (Swertia chirata: 21147), (Caesalpinia bonduc:21148), (Citrullus colocynthis:21149), (Bunium persicum:21150), (Cuminum cyminum; 21151). For extract preparation, 100 grams of each plant were washed, shade dried at 25+2°C and powdered. The powders were decocted in boiling water (1:9) for 30 min, cooled and filtered. The filtrates were dried, mixed in equal ratio to prepare polyherbal formulation and stored at 2-8°C.

Qualitative phytochemical analysis: Phytochemical analysis of polyherbal formulation was performed for detection of alkaloids (Salehi-Surmaghi *et al.*, 1992), carbohydrates (Gupta *et al.*, 2013), tannins (Segelman *et al.*, 1969), phenolics and flavanoids by FeCl₃ test described by Mace (1963) and Raman (2006). Keller-kiliani test was performed for detection of glycosides (Ajaiyeobu *et al.*, 2002), stain test for fixed oil (Shabi *et al.*, 2014), frothing test for saponins (Kumar *et al.*, 2009) and Salkowski test for terpenoids (Evans, 1997).

Research study protocol and experimental induction of diabetes: We housed ninety (90) healthy young albino rats (180-200 g) in an animal facility at Institute of Pharmacy, Physiology and Pharmacology, University of Agriculture, Faisalabad. Rats were divided into 6 groups, each group having 15 rats. Group I was control, group II was diabetic and group III was +ve control. Group IV, V and VI were treated I, treated II and treated III. The experiment was conducted according to approved methods of Graduate Studies Research Board. Rats of group II, III, IV, V and VI were induced diabetes by using single intraperitoneal injection of alloxan monohydrate (150 mg/kg BW) dissolved in normal saline.

Administration of polyherbal formulation to alloxanized hyperglycemic rats and blood sampling: After seven days of alloxan induction, group III was treated with 10 mg/kg of glibenclamide and group IV, V and VI were administered 200, 400 and 600 mg/kg of polyherbal formulation extract respectively through intragastric tube upto 8 weeks of experiment. After that, the rats were slaughtered by cervical dislocation and blood samples were collected to separate serum for further analysis. Their pancreata were dissected out and snapfrozen in liquid nitrogen for gene expression analysis.

Biochemical analysis: Serum glucose, serum insulin and Hb1Ac levels were determined by using commercially available kits. Hepatic glucokinase level was assayed spectrophotometrically by measuring absorbance of reaction cocktail consisting of tissue homogenate and glucose-6-phosphate dehydrogenase assay system at 340nm (Goward *et al.*, 1986).

Lipid Profile: Serum total cholesterol, triglycerides and high-density lipid (HDL) levels were determined by using commercially available kits (Dia Sys Diagnostic Systems USA reagent kit method). Low density lipoproteins (LDL) and very low-density lipoproteins (VLDL) concentrations were calculated by using following formulae (Friedewald *et al.*, 1972).

LDL-cholesterol = Cholesterol - $\left[\frac{\text{Triglyceride}}{5} + \text{HDL-Cholesterol}\right]$

 $VLDL\text{-cholesterol} = [\frac{\text{Triglycerides}}{5}]$

Gene expression analysis: RNA isolation was performed by using TRIzol method (ThermoFisher Scientific, Waltham, Massachusetts, USA) (Liu and Patel, 1995), quantified on nanodrop and subjected to cDNA synthesis with equal RNA concentration in each sample using the RevertAid cDNA synthesis kit (ThermoFisher Scientific). qRT-PCR was performed using Maxima SYBR Green/ ROX qRT-PCR Master Mix (ThermoFisher Scientific). Expression of different genes involved in insulin signaling pathway (Pdx-1, IGF-1, Ins-1) was analyzed. The primer sequences of genes are given in Table 1. The cDNA was denatured for 15 seconds at 95°C for all genes. Then primers were annealed at 52°C for 25 seconds and extension time was 20 seconds at 72°C for 40 cycles. All these steps were accompanied by 95°C denaturation for 10 minutes at the start of a single cycle. Expression levels of genes were normalized to β -actin. The 2*(- $\Delta\Delta$ ct) method was used to analyze qRT-PCR data.

RESULTS

Phytochemical analysis of polyherbal formulation (**Qualitative analysis**): Crude aqueous extract of polyherbal formulation was subjected to qualitative analysis. The results of various chemical tests for the detection of phytochemicals are summarized in Table 2. Results have indicated the presence of alkaloids, phenolic compounds, flavonoids, glycosides, saponins, carbohydrates and terpenoids in polyherbal formulation.

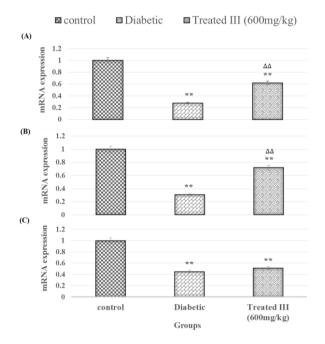


Fig. 1: Rat's pancreatic gene expression profile: (A) mRNA expression of Pdx-1 in control, diabetic and polyherbal formulation treated rat's pancreata (n=3) (B) mRNA expression of Ins-1 in control, diabetic and polyherbal formulation treated rat's pancreata (n=3) (C) mRNA expression of IGF-1 in control, diabetic and polyherbal formulation treated rat's pancreata (n=3). (**P≤0.01 shows significant difference between control and other groups. $^{\Delta\Delta}P \leq 0.01$ indicates significant difference between diabetic and polyherbal formulation treated group III).

Table I: Name and source of oligonucleotides (Primers). All oligonucleotides listed below were supplied by Invitrogen, Karlsruhe, Germany

| Indeleonaes listed | below were supplied by invite ogen, realisitune, German |
|--------------------|---|
| Beta actin F | 5'- CGAGTACAACCTTCTTGCAGC -3' |
| Beta actin R | 5'- TATCGTCATCCATGGCGAACTG -3' |
| Pdx1 F | 5'- TCCCGAATGGAACCGAGACT -3' |
| Pdx I R | 5′- TTCATCCACGGGAAAGGGAG -3′ |
| Ins I F | 5'- AGGCTCTGTACCTGGTGTGTG -3' |
| Ins I R | 5'- CGGGTCCTCCACTTCACGAC -3' |
| IGFI F | 5′- CGTACCAAAATGAGCGCACC -3′ |
| IGFI R | 5′- CAAGCAGAGTGCCAGGTAGA -3′ |
| | |

Table 2: Qualitative analysis of polyherbal formulation

| Phytochemical | Present/Absent |
|---------------|----------------|
| | TresenuAbsent |
| Phenolics | + |
| Alkaloids | + |
| Flavanoids | + |
| Saponins | + |
| Glycosides | + |
| Fixed oils | + |
| Tannins | - |
| Carbohydrates | + |
| Terpenoids | + |

(+)= Present, (-)=Absent).

Biochemical analysis

Effect of polyherbal formulation on serum glucose, insulin and HbA1C: Serum glucose and glycosylated hemoglobin (HbA1c) levels were significantly (P≤0.01) increased in hyperglycemic rats. Glibenclamide produced significant reduction in serum glucose and HbA1C concentrations. However, treatment of diabetic rats with polyherbal formulation at 200, 400 and 600mg/kg upto 8 weeks significantly ($P \le 0.01$) lowered the serum glucose and glycosylated hemoglobin levels in treated groups I. II and III respectively compared to diabetic group. Diabetic rats showed significant (P<0.01) decrease in serum insulin level when compared to control group after induction of diabetes. This significant decrease was successfully reversed in the diabetic rats upon treatment with graded doses of polyherbal formulation extract for 8 weeks in dose dependent manner (Table 3).

Effect of polyherbal formulation on glucokinase enzyme: A significant decrease in glycolytic enzyme activity (glucokinase) was observed in diabetic, +ve control and polyherbal formulation treated groups I, II and III after administration of alloxan monohydrate compared to control group. Polyherbal formulation treatment of diabetic rats of treated groups I, II and III for 8 weeks showed significant (P \leq 0.01) elevation in hepatic glucokinase activity when compared to diabetic group (Table 3).

Effect of polyherbal formulation of lipid profile of alloxanized hyperglycemic rats: Significant change in serum total cholesterol, triglycerides, HDL-cholesterol, LDL and VLDL levels were observed in alloxanized hyperglycemic rats. Polyherbal formulation supplementation (200, 400 and 600 mg/kg) to diabetic rats of treated groups I, II and III up to 8 weeks showed significant (P≤0.01) decrease in serum total cholesterol, triglycerides, LDL and VLDL levels where as valuable elevation in HDL-cholesterol level was observed compared to diabetic group in dose dependent manner (Table 4). Results have also exhibited that highest dose (600 mg/kg) of polyherbal formulation was more effective in producing antihyperlipidemic effect compared to glibenclamide.

qRT-PCR based gene expression analysis (Effect on insulin signaling pathway): The effect of polyherbal formulation on mRNA expression of Ins-1, Pdx-1 and IGF-1 genes in control, diabetic and highest dose (600 mg/kg) of polyherbal formulation treated group has been shown in Fig. 1a-c. As the highest dose of herbal mixture extract was proved to be more effective compared to other doses according to biochemical results, that's why only highest dose treated group was compared with control and diabetic group in gene expression analysis. From this analysis, it was found that Ins-1, Pdx-1 and IGF-1 genes expressions were significantly (P≤0.01) reduced in diabetic and polyherbal formulation treated group after induction of diabetes compared to control group. Polyherbal formulation treatment of hyperglycemic rats significantly up regulated the mRNA expression of Ins-1 and Pdx-1 genes compared to diabetic group. However, mRNA expression of IGF-1 differ non-significantly between diabetic and 600mg/kg of polyherbal formulation treated group.

Table 3: Effect of graded doses of polyherbal formulation and glibenclamide on serum glucose, insulin, HbA1c and glucokinase level of alloxan induced diabetic rats

| Treatment | Control | Diabetic | +Ve control | Treated I | Treated II | Treated III |
|---|--------------|--------------|--------------|--------------|--------------|--------------|
| Serum Glucose (mg/dL) | 109.80±4.28D | 518.00±5.31A | 138.20±2.65C | 233.60±2.62B | 224.20±3.38B | 143.1±3.07BC |
| Serum Insulin (mg/dL) | 17.95±0.69A | 6.58±0.75D | 17.65±0.73B | 12.23±0.71C | 14.65±0.67BC | 15.78±0.64BC |
| HbAIc (%) | 5.87±0.03E | 13.92±0.07A | 6.14±0.02D | 7.70±0.03B | 6.93±0.03C | 6.84±0.03D |
| Glucokinase (µmolG6PO4/min/mg proteins) | 198.6±1.30A | 111.2±3.24D | 181.6±1.23B | 154.0±2.03C | 164.4±1.67BC | 176.2±1.84B |

One-way ANOVA followed by DMR test. Values (Mean+SE) with different superscripts within a row differ significantly ($P \leq 0.01$).

Table 4: Effect of graded doses of polyherbal formulation and glibenclamide on serum lipid profile of alloxan induced diabetic rats after 8 week treatment

| Parameters | Control | Diabetic | +Ve control | Treated I | Treated II | Treated III |
|---------------------------|--------------|--------------|---------------|--------------|---------------|--------------|
| Total Cholesterol (mg/dL) | 120.20±3.62E | 266.80±5.48A | 140.20±3.25CD | 142.00±2.47B | 128.20±3.06BC | 111.00±2.70D |
| Triglycerides (mg/dL) | 69.20±1.28E | 164.60±3.03A | 91.00±1.95C | 110.40±2.27B | 109.40±2.69B | 81.80±1.32D |
| HDL-C (mg/dL) | 40.60±0.93A | 17.80±1.83E | 32.40±0.68CD | 27.80±0.58D | 31.40±0.51CD | 34.20±0.73B |
| LDL (mg/dL) | 65.12±1.60E | 216.04±7.51A | 89.60±0.51CD | 92.12±0.73B | 74.91±0.42BC | 60.44±1.03D |
| VLDL (g/dL) | 13.86±0.37E | 32.96±0.83A | 18.2±0.13C | 22.08±0.27B | 21.89±0.34B | 16.36±0.20D |

One-way ANOVA followed by DMR test. Values (Mean+SE) with different superscripts within a row differ significantly (P≤0.01).

DISCUSSION

Herbal therapies for treatment of diabetes are being used from decades both in insulin dependent and noninsulin dependent diabetic patients. Herbal products contain more than one active constituent compared to allopathetic treatment. These phyto constituents having different structure but with same curative potential act in synergistic way for treatment of different diseases (Mukherjee et al., 2006). Results of phytochemical analysis of polyherbal formulation indicated the presence of alkaloids, flavonoids, phenols, glycosides and saponins in herbal mixture extract. These phytochemicals (alkaloids, saponins, polyphenols, flavonoids) may influence glucose metabolism by different mechanisms like stimulation of insulin release from the pancreatic β cells, activation of insulin receptors, glucose uptake in the insulin sensitive cells, increasing glucokinase activity and controlling glucose release from liver (Ramadan et al., 2017).

The results have shown that alloxan monohydrate significantly (P \leq 0.01) raised serum glucose, glycosylated hemoglobin (HbA1c) and decreased serum insulin and glucokinase levels in all groups except control group. Interestingly, treatment of diabetic rats with polyherbal formulation resulted in significant glycemic control by lowering serum glucose and raising insulin secretion. Possible mechanism of action of antihyperglycemic rats may be due to increased insulin discharge from existing β cells as well as increased transfer of glucose into peripheral tissues. qRT-PCR analysis also confirmed upregulation of Pdx-1 and Ins-1 genes which are directly related with pancreatic β cells regeneration and insulin secretion (Jie *et al.*, 2016).

Glucokinase (hexokinase IV) is present in the liver and pancreas of most vertebrates. It represents 95% of hexokinase activity in liver (Kawai *et al.*, 2005). In pancreatic β cells, glucokinase serves as a glucose sensor to modify insulin discharge. Its level is controlled by insulin, not glucose-6-phosphate. Due to this property, liver stores glucose-6-phosphate and converts it into glycogen for later use. Mutations in glucokinase have been associated with diabetes mellitus (Martin *et al.*, 2008). Treatment of diabetic rats with graded doses of polyherbal formulation significantly (P≤0.01) increased hepatic glucokinase activity resulting in increased utilization of glucose and formation of liver glycogen.

Hyperlipidaemia is a common complication of diabetes mellitus. The present study results identified significant (P<0.01) change in lipid profile. Treatment of diabetic rats with different doses of polyherbal formulation extract resulted in gradual decrease in serum cholesterol, triglycerides, low-density lipoproteins (LDL) and very low-density lipoproteins (VLDL) levels with increase in HDL level. The lipid-lowering trend of aqueous polyherbal formulation extract may be due to the presence of flavonoids which are reported to lower the levels of cholesterol and triglycerides (Hossain et al., 2016). HDL is known as good cholesterol and scavenger because it carries LDL away from arteries. Various research studies have reported that increase in HDLcholesterol is linked with decline in coronary disease (Momo et al., 2006).

qRT-PCR based gene expression analysis has been performed to analyze mRNA expression of different pancreatic genes involved in insulin signaling pathway like Pdx-1, Ins-1 and IGF-1. Results have shown that alloxan monohydrate significantly (P≤0.01) down regulated the Ins-1, Pdx-1 and IGF-1 mRNA expressions. The IGF-1 is a growth factor and has insulin-like effects on cell metabolism (Siddique and Awan, 2016). Pancreatic and duodenal homeobox factor-1 (Pdx-1) is expressed in the pancreas and duodenum and plays crucial role in pancreatic β-cells differentiation/regeneration (Kubo et al., 2011). Results of our research work have shown that there appeared non-significant (P>0.05) difference in expression level of IGF-1 between diabetic and polyherbal formulation (600 mg/kg) treated groups. However, mRNA expression of Pdx-1 and Ins-1 were significantly ($P \le 0.01$) reversed by the polyherbal formulation treatment. This finding further strengthens the role of polyherbal formulation in reversing hyperglycemic and diabetic conditions.

Conclusions: In conclusion, our research work has revealed that polyherbal formulation is an effective antidiabetic agent as it can reduce hyperglycemia, improve lipid profile, prevent pancreatic β cells apoptosis and ameliorate insulin secretary capacity. However further research is required to elucidate its mechanism of action. **Acknowledgements:** We would like to thank Director, Institute of Pharmacy, Physiology and Pharmacology for providing facilities for the conduct of experiment. We are also thankful to institute of microbiology for providing qRT-PCR facility for gene expression analysis.

Authors contribution: This manuscript is based on PhD thesis of the 1st author. AI and BA contributed to design the whole experiment. TK and AI analyzed the samples. FM statistically analyzed the data. All authors were involved in discussing the contents of the manuscript and agreed to publication.

REFERENCES

- Abbas A, Iqbal Z, Abbas RZ, et al., 2017a. Immunomodulatory activity of Pinus radiata extract against coccidiosis in broiler chicken. Pak Vet J 37:145-9.
- Abbas A, Iqbal Z, Abbas RZ, et al., 2017b. In vivo anticoccidial effects of Beta vulgaris (sugar beet) in broiler chickens. Microb Path 111:139-44.
- Ajaiyeobu EO, 2002. Phytochemical and antibacterial activity of Parkia biglobosa and Parkia bicolor leaf extracts. Afr J Biomed Res 5:125-9.
- Almalki WH, Arafa ESA, Abdallah AY, et al., 2016. Zinc chloride protects against streptozotocin-induced diabetic nephropathy in rats. Pharmacol Pharm 7:331-42.
- Dey L, Attele AS and Yuan CS, 2002. Alternative therapies for type 2 diabetes. Altern Med Rev 7:45-58.
- El-Abhar HS and Schaalan MF, 2014. Phytotherapy in diabetes: review on potential mechanistic perspectives. World J Diabetes 5:176-97.
- Evans WC, 1997. In: Pharmacology: Harcourt Brace and Company, Asia, Singapore, pp:226-8.
- Friedewald WT, Levy RI and Fredrickson DS, 1972. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem 18:499-502.
- Ghorbani A, 2014. Clinical and experimental studies on polyherbal formulations for diabetes: current status and future prospective. J Integr Med 12:336-45.
- Goward CR, Hartwell R, Atkinson T, et al., 1986. The purification and characterization of glucokinase from the thermophile bacillus stearothermophilus. Biochem J 15:415-20.
- Gupta M, Thakur S, Sharma A, et al., 2013. Qualitative and quantitative analysis of phytochemicals and pharmacological value of some dye yielding medicinal plants. Orient J Chem 29:475-81.
- Hatware K and Annapurna A, 2014. The effect of quercetin on blood glucose levels of normal and streptozotocin induced diabetic (type i & type ii) rats. Int J Pharmaceut Chem Biol Sci 4:613-9.
- Hassan KU, Khalique A, Pasha TN, et al., 2017. Influence of Moringa oleifera decorticated seed meal on broiler performance and immunity. Pak Vet J 37: 47-50.
- Hossain MK, Dayem AA, Han J, et al., 2016. Molecular mechanisms of the anti-obesity and anti-diabetic properties of flavonoids. Int J Mol Sci 17:569.
- Hossen MJ, Uddin MB, Ahmed SSU, et al., 2016. Traditional medicine/plants for the treatment of reproductive disorders in Asia nations. Pak Vet J 36: 127-133.
- Jie Guo J, Wanga J, Song S, et al., 2016. Sphallerocarpus gracilis polysaccharide protects pancreatic β cells viaregulation of the bax/bcl-2, caspase-3, pdx-1 and insulin signallingpathways. Int J Biol Macromol 93:829-36.

- Kawai S, Mukai T, Mori S, et al., 2005. Hypothesis: structures, evolution, and ancestor of glucose kinases in the hexokinase family. J Biosci Bioeng 99:320-30.
- Kubo A, Stull R, Takeuchi M, et al., 2011. Pdx1 and Ngn3 overexpression enhances pancreatic differentiation of mouse ES cell-derived endoderm population. PLoS ONE 6:24058.
- Kumar A, Ilavarasan R, Jayachandran T, et al., 2009. Phytochemical investigation on a tropical plant. Pak J Nutr 8:83-5.
- Liaqat I, Pervaiz Q, Bukhsh SJ, et al., 2016. Investigation of bactericidal effects of medicinal plant extracts on clinical isolates and monitoring their biofilm forming potential. Pak Vet | 36: 159-164.
- Liu JL and Patel YC, 1995. Glucocorticoids inhibit somatostatin gene expression through accelerated degradation of somatostatin messenger ribonucleic acid in human thyroid medullary carcinoma (TT) cells. Endocrinol 136:2389-96.
- Mace SL, 1963. Anaerobic bacteriology for clinical laboratories. Pharmacognosy 23:89-91.
- Mannan A, Rupa BA, Azam NK, et al., 2014. A quick review on antidiabetic plants and action of phytochemicals. Inter J Adv Res 2:227.
- Martin D, Bellanné-Chantelot C, Deschamps I, et al., 2008. Long-term follow-up of oral glucose tolerance test-derived glucose tolerance and insulin secretion and insulin sensitivity indexes in subjects with glucokinase mutations (MODY2). Diabetes Care 31:1321-3.
- Momo CEN, Julius EO and Dagobert T, 2006. Antidiabetic and hypolipidemic effects of *Laportea ovalifolia* (URTICACEAE) in alloxan induced diabetic rats. African J Trad Comp Alter Med 3:36-43.
- Mukherjee PK, Mukherjee K and Maiti K, 2006. Leads from Indian medicinal plants with hypoglycemic potentials. J Ethnopharmacol 106:1-28.
- Paoli A, Bosco G, Camporesi EM, et al., 2015. Ketosis, ketogenic diet and food intake control: a complex relationship. Front Psychol 6:27.
- Piya MK, Tahrani AA and Barnett AH, 2010. Emerging treatment options for type 2 diabetes. J Clin Diabetes Br Pharmacol 70:631-44.
- Ramadan BK, Schaalan MF and Tolba AM, 2017. Hypoglycemic and pancreatic protective effects of Portulaca oleracea extract in alloxan induced diabetic rats. BMC Comlt and Alt Med 17:37.
- Raman N, 2006. Phytochemical Methods: New Indian Publishing Agencies, New Delhi, pp:19.
- Riserus U, Willett WC and Hu FB, 2009. Dietary fats and prevention of type 2 diabetes. Prog Lipid Res 48:44-51.
- Salehi-Surmaghi MH, Aynehchi Y, Amin GH, et al., 1992. Survey of Iranian plants for saponins, alkaloids, flavonoids and tannins. IV. DARU J Pharm Sci 2:1-11.
- Segelman AB, Farnsworth NR and Quimby MD, 1969. False negative saponins test results induced by the presence of tannins. Lloydia, 32:52-8.
- Shabi RR, Kumari V and Chitarasu, 2014. Qualitative phytochemical characterization of thorn extracts of *Canthium parviflorum* Lam. Int Res J Pharm App Sci 4:13-7.
- Si YL, Zhao YL and Hao HJ, 2011. MSCs: biological characteristics, clinical applications and their outstanding concerns. Ageing Res Rev 10:93-103.
- Siddique T and Awan FR, 2016. Effects of Reg3 delta bioactive peptide on blood glucose levels and pancreatic gene expression in an alloxan-Induced mouse model of diabetes. Can J Diabetes 40:198-203.
- Tahir M, Rasheed MA, Niaz Q, et al., 2017. Evaluation of antibacterial effect of *Gymnema sylvestre* R.Br. species cultivated in Pakistan. Pak Vet J 37: 245-250.